

# Microbial monooxygenases

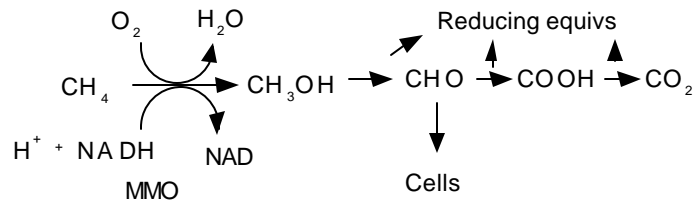
Stuart E Strand

## Dechlorination of chlorinated aliphatic hydrocarbons by methane-oxidizers, methanotrophs

Methanotrophs are obligate methane oxidizers, obligate aerobes, important part of carbon cycle, completing oxidation of methane to CO<sub>2</sub>.

Widely distributed, particularly at the interface between aerobic and anaerobic environments from which methane diffuses to the atmosphere.

Methane monooxygenase first step in methane oxidation to CO<sub>2</sub>



Requires oxygen and energy (NADH). NADH is produced by the oxidation of methanol, formaldehyde and formate.

Methane monooxygenase comes in two varieties: soluble and membrane bound. The soluble MMO (sMMO) is very nonspecific and is present primarily in Type II methanotrophs, where it is induced by low copper conditions enzyme (<50 µg/L). The membrane bound or particulate MMO (pMMO) is constitutive, but limited by the availability of copper. It is more specific for methane and oxidizes fewer xenobiotics

Methanotrophs are classified by the pathway used to assimilate carbon from formaldehyde, their internal membrane structure, G+C content of DNA, and metabolism:

Type	Representative species	C assimilative pathway	G+C%	Metabolism	Internal membranes	Type of MMO
I	<i>Methylomonas albus</i> BG8	hexulose monophosphate pathway	50-54	incomplete TCA, unable to fix N <sub>2</sub>	bundles or stacks	Mostly pMMO
II	<i>Methylosinus trichosporium</i> OB3b	serine pathway	58-66	complete TCA, able to fix N <sub>2</sub>	paired peripheral membranes	Both pMMO & sMMO

The soluble sMMO attacks trichloroethylene; 1,1,1-trichloroethane; mono & dichloroethane, methane and ethene; chloroform; others:

### Environmentally Significant Compounds

#### Oxidized by Methanotrophs

Compound	Product (pure culture)
Chloromethane	Formaldehyde
Dichloromethane	Carbon monoxide
Trichloromethane(Chloroform)	Carbon dioxide
Bromomethane	Formaldehyde
Benzene	Phenol & Hydroquinone
Toluene	<i>p</i> -Cresol, Benzyl alcohol, Benzoate
Styrene	Styrene oxide & Hydroxystyrene
m-Cresol	Hydroxybenzaldehydes
o-Cresol	5-Methyl-1,3-benzenediol
m-Chlorotoluene	Benzyl alcohols
Naphthalene	Naphthols
1- and 2- Methylnaphthalene	ND
Vinyl chloride	ND
1,2 & 1,1-dichloroethylene	ND
Trichloroethylene	Dichloroacetic acid & TCE Diol
1,1- & 1,2-dichloroethane	ND
1,1,1- & 1,1,2-trichloroethane	ND

The oxidation of these compounds provides no benefit for the methanotrophs and drains the organism of energy (NADH); thus it is **cometabolism**

There are 3 components to the sMMO: the hydroxylase, the reductase, and a component B. The reactions involving methane occur at the hydroxylase. The active site has a unique hydroxo-bridged dinuclear iron center. The diiron is first reduced to ferrous, which then reacts with O<sub>2</sub> to yield water and a di-Fe[IV]=O species, which pulls an H from CH<sub>4</sub> to form a CH<sub>3</sub>\* radical and diiron cluster hydroxyl radical. Recombination forms methanol. Comp B and the reductase feed NADH electron transfer and regulate the reaction by forming complexes that alter MMO activities. (Lipscomb, J. D. (1994). "Biochemistry of the soluble methane monooxygenase." Annual Review of Microbiology **48**(1994): 371-399)

Four types of inhibitory effects on the methanotrophs:

1. Depletion of energy (NADH)
2. Competition with methane for the MMO site, competitive inhibition with the following kinetics:

$$\frac{dS_t}{dt} = r_{tm} \frac{S_t/K_t}{1 + S_s/K_s + S_t/K_t}$$

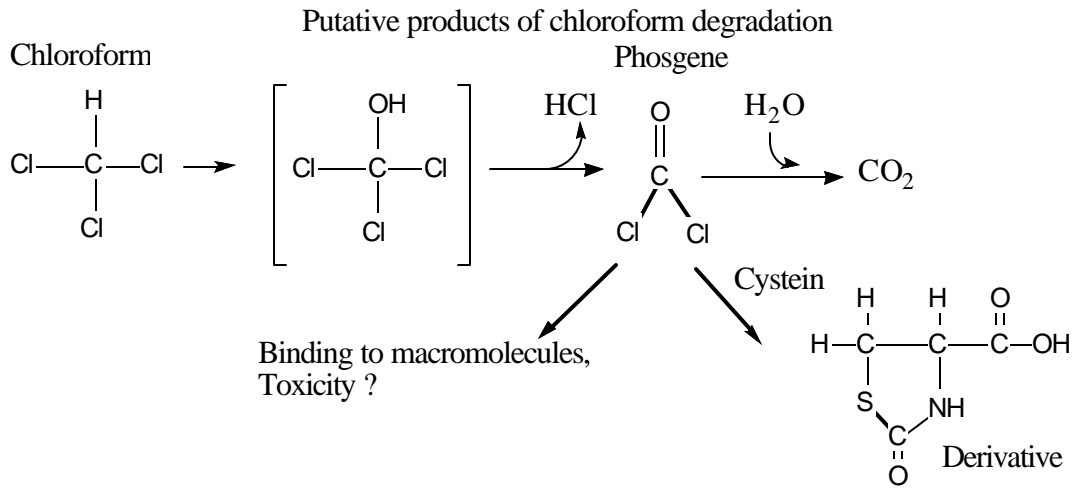
S<sub>t</sub> is the concentration of the toxic compound, K<sub>t</sub> is the affinity constant for the toxic compound,

S<sub>s</sub> is the concentration of the growth substrate, methane, K<sub>s</sub> is the affinity constant for methane.

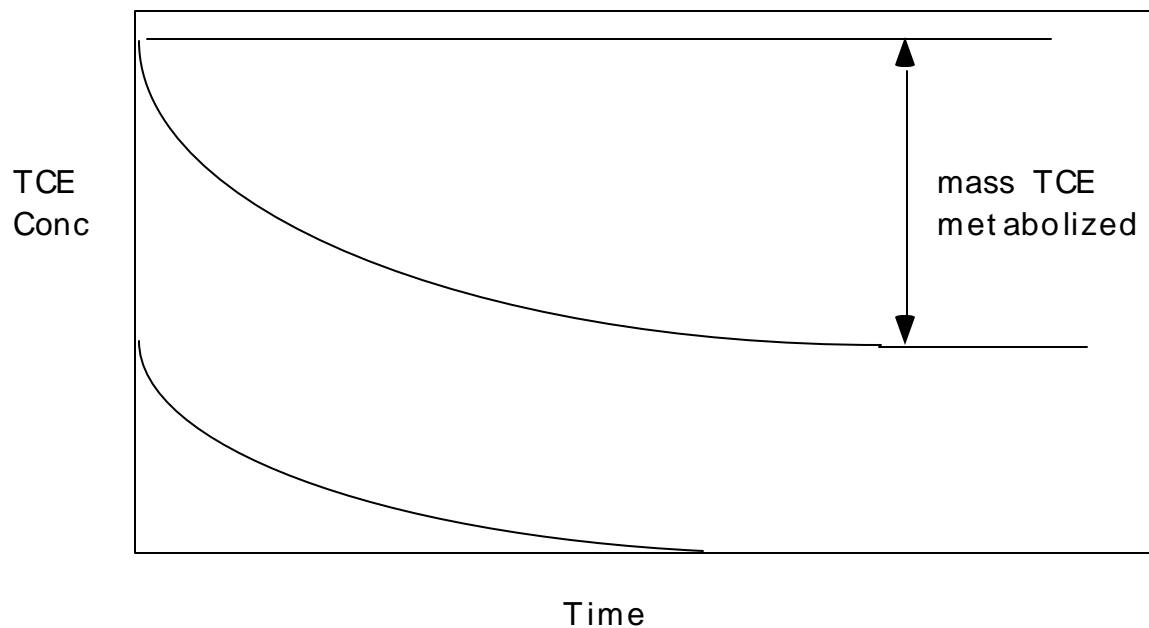
This expression degenerates to the Michaelis-Menten rate expression if S<sub>s</sub> is zero

3. Toxic effects of the chlorinated solvents on membranes and metabolism
4. Toxic effects of the metabolites, Particularly in the case of TCE and Chloroform. These metabolites are the same as those that cause damage in the mammalian transformations by cytochrome P450

Toxic metabolites:



Metabolite toxicity effects detectable by course of CF transformation.



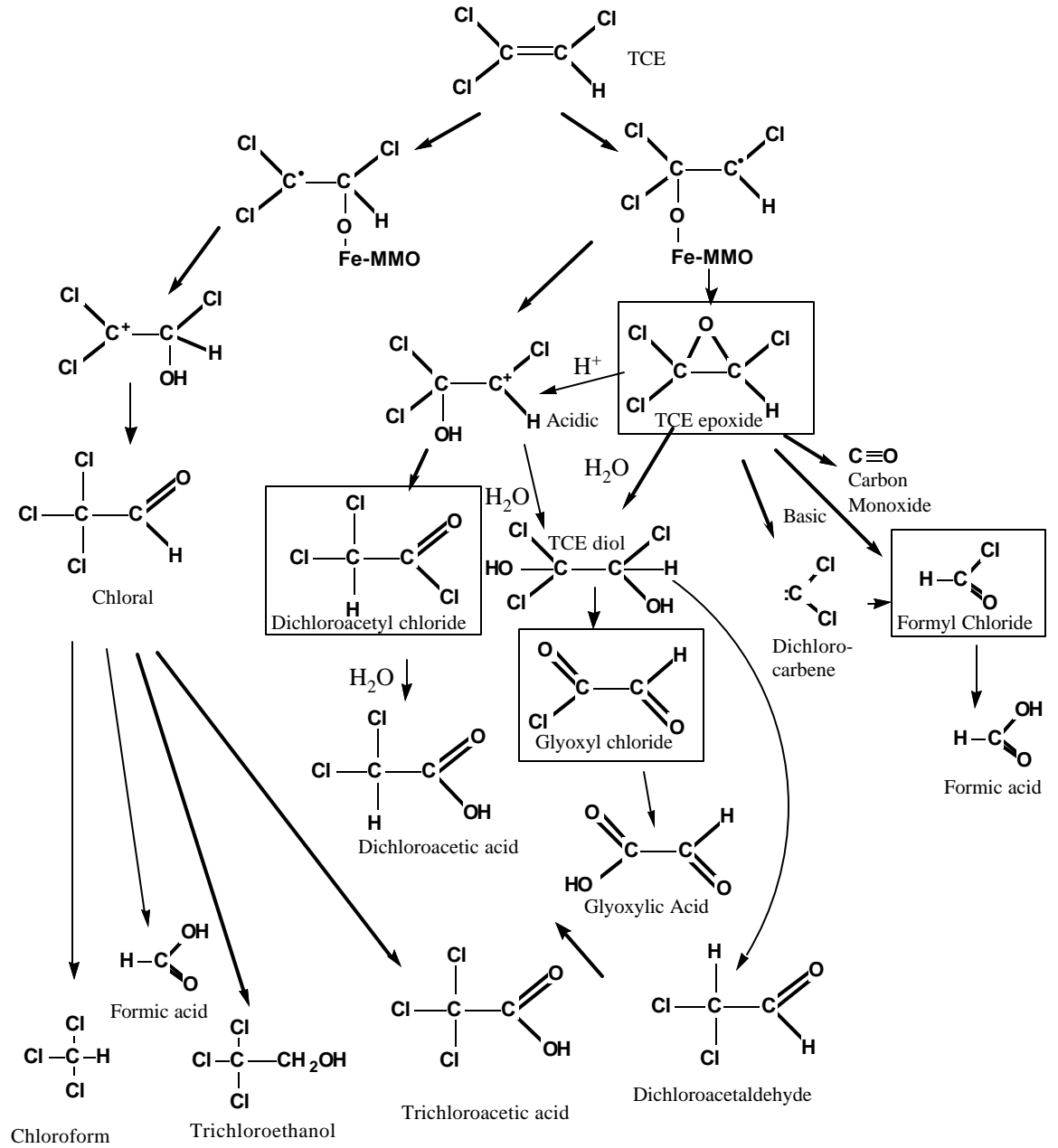
At the end of the time for the higher TCE concentration, the cells will be inactivated, unable to oxidize either TCE or their substrate.

Quantify toxicity effect using transformation capacity:

$$T_C = (\text{mass TCE metabolized})/(\text{biomass inactivated})$$

## Putative products of TCE degradation

The products most likely to covalently bind enzymes resulting in cell death are shown in boxes.



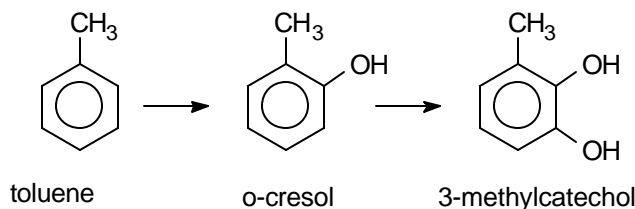
### Propane and ethylene oxidizers

Note that propane and ethylene oxidizers also have monooxygenases with broad nonspecificity. Propane oxidizers especially may be useful for oxidation of 1,1,1-trichloroethane and chloroform in addition to TCE (Keenan, J. E., S. E. Strand and H. D. Stensel (1993). Degradation kinetics of Chlorinated Solvents by a Propane-Oxidizing Enrichment Culture. Bioremediation of Chlorinated and Polycyclic Aromatic Hydrocarbon Compounds. R. E. Hinchee, A. Leeson, L. Semprini and S. K. Ong. Boca Rton, Lewis Publishers: 1-13).

### The ammonia monooxygenase enzyme of nitrification

The monooxygenase of nitrifying organisms is similar to that of the methanotrophs. The AMO oxidizes methane and most of the compounds oxidized by the methanotrophs, but at lower rates. TCE and Chloroform produce toxic products. Ammonium is more soluble than methane, but nitrate is a pollutant. Since MMO and AMO are inhibited by the same inhibitors (esp acetylene), their environmental activities may be hard to distinguish.

### Toluene monooxygenases



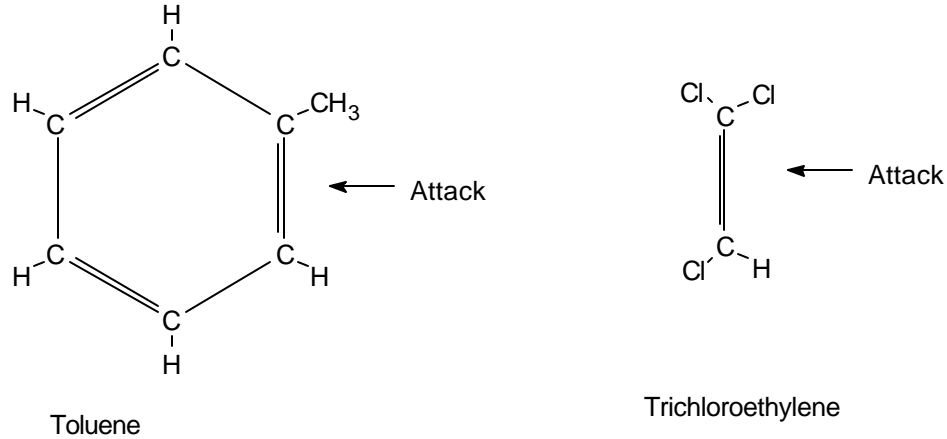
**Toluene 2-monooxygenase** of *Burkholderia* (formerly *Pseudomonas*) *cepacia* G4. High rates of TCE degradation, with little intermediate toxicity (Newman, L. M. and L. P. Wackett (1995). "Purification and Characterization Of Toluene 2-Monooxygenase From Burkholderia Cepacia G4." Biochemistry **34**(43): 14066-14076.)

**Toluene 2,3-dioxygenase** of *Pseudomonas putida* F1 acts as a monooxygenase with TCE. High rate, high intermediate toxicity. (Jahng, D. J. and T. K. Wood (1994). "Trichloroethylene and chloroform degradation by a recombinant pseudomonad expressing soluble methane monooxygenase from Methylosinus trichosporium OB3b." Applied and Environmental Microbiology **60**(7): 2473-2482; Spain, J. C., G. J. Zylstra, C. K. Blake and D. T. Gibson (1989). "Monohydroxylation of phenol and 2,5-dichlorophenol by toluene dioxygenase in *Pseudomonas putida* F1." Appl. Environ. Microbiol. **55**: 2648-2652; Wackett, L. P., L. D. Kwart and D. T. Gibson (1988). "Benzylic monooxygenation by toluene dioxygenase from *Pseudomonas putida* ." Biochemistry **27**: 1360-1367)

Also **toluene-4-monooxygenase** of *Pseudomonas mendocina* KR1, a poor TCE degrader (Yen, K. M. and M. R. Karl (1992). "Identification of a new gene, tmoF, in the *Pseudomonas*

*mendocina* KR1 gene cluster encoding toluene-4-monooxygenase.” *J. Bacteriol* **174**(22): 7253-7261.)

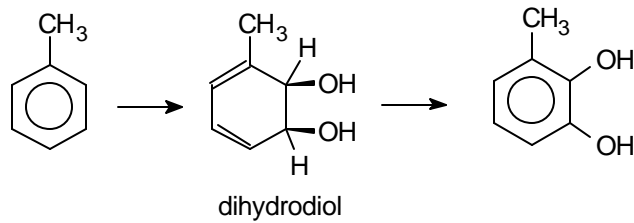
Aromatic oxygenases, Oxidize chlorinated ethylenes, but not chloroform or 111-TCA



### Compare

#### Toluene dioxygenase

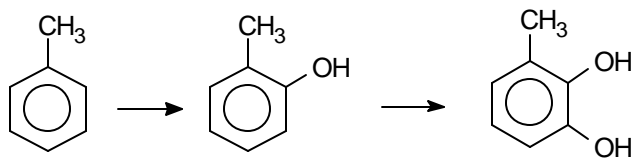
*Pseudomonas putida* F1, grows on toluene



toluene dioxygenase

#### Toluene ortho monooxygenase

*Pseudomonas cepacia* G4, grows on phenol or toluene



toluene ortho monooxygenase

### Fungal P450s

Fungal and bacterial cytochrome P450s are the enzymes used in the initial oxidation of alkanes and aromatics of petroleum. Some yeast (esp. *Saccharomyces cerevisiae*) use P450s in sterol synthesis, but they also have nonspecific cometabolic effects.

The P450 also can reduce fully halogenated compounds under anaerobic conditions: carbon tetrachloride → chloroform, but only in mammalian and plant metabolism

### Bibliography

#### Cometabolism and methanotrophs

- Bauer JE, Capone DG (1988) Effects of co-occurring aromatic hydrocarbons on degradation of individual polycyclic aromatic hydrocarbons in marine sediment slurries. **Appl. Environ. Microbiol.** 54, 1649-1655.
- Apajalahti, J.H.A., Salkinoja-Salonen, M.S. (1987) Dechlorination and *para*-hydroxylation of polychlorinated phenols by *Rhodococcus chlorophenolicus*. **Jour. Bacteriol.** 169, 2, 675-681.
- Dagley, S. (1985) Microbial metabolism of aromatic compounds. in **Comprehensive Biotechnology**. (Moo-Young, M., Bull, A. T., Dalton, H., eds.) 1, Pergamon Press, Oxford, 483-505.
- Guerin WF, Jones GE (1988) Two-stage mineralization of phenanthrene by estuarine enrichment cultures. **Appl. Environ. Microbiol.** 54, 929-936.
- Kim, C. J., Maier, W. J. (1986) Acclimation and biodegradation of chlorinated organic compounds in the presence of alternate substrates. **Appl. Environ. Microbiol.** 58, 157-163.
- Little, C. D., Palumbo, A. V., Herbes, S. E., Lidstrom, M. E., Tyndall, R. L., Gilmer, P. J. (1988) Trichloroethylene biodegradation by a methane-oxidizing bacterium. **Appl. Environ. Microbiol.** 54, 951-956.

Reineke, W., Knackmuss, H.-J. (1984) Microbial metabolism of haloaromatics: isolation and properties of a chlorobenzene-degrading bacterium. **Appl. Environ. Microbiol.** 47, 395-402.

Spain JC, Van Veld PA (1988) Adaptation of natural microbial communities to degradation of xenobiotic compounds: Effects of concentration, exposure time, inoculum, and chemical structure. **Appl. Environ. Microbiol.** 45, 428-435.

Stirling, D. I., Dalton, H. (1979) The fortuitous oxidation and cometabolism of various carbon compounds by whole-cell suspensions of *Methylococcus capsulatus* (Bath). **FEMS Microbiol. Letters** 5, 315-318.

Green J, Dalton H (1989) Substrate specificity of soluble methane monooxygenase. mechanistic implications. *J. Biol. Chem.* 264, , 17698-17703.

Fox BG, Borneman JG, Wackett LP, Lipscomb JD (1990) Haloalkene oxidation by the soluble methane monooxygenase from *Methylosinus trichosporium* OB3b: Mechanistic and environmental implications. *Biochemistry* 29, 6419-6427.

#### The nitrifiers

Hyman MR, Wood PM (1983) Methane oxidation by *Nitrosomonas europaea*. *Biochem. J.* 212, 31-37

Hyman, M. R., Sansome-Smith, A. W., Shears, J. H., Wood, P. M. (1985) A kinetic study of benzene oxidation to phenol by whole cells of *Nitrosomonas europaea* and evidence for the further oxidation of phenol to hydroquinone. *Arch. Microbiol.*, 143, 302-306

Arciero D, Vannelli T, Logan M, Hooper AB (1989) Degradation of trichloroethylene by the ammonia-oxidizing bacterium *Nitrosomonas europaea*. *Biochem. Biophys. Res. Comm.* 159, 640-643

Vannelli T, Logan M, Arciero D M, Hooper A B (1990) Degradation of halogenated aliphatic compounds by the ammonia-oxidizing bacterium *Nitrosomonas europaea*. *Applied and Environmental Microbiology*, 56, 1169-1171

## Fungal P450s

Azari, M. R., Wiseman, A. (1982) Purification and characterization of the cytochrome P-448 component of a benzo(a)pyrene hydroxylase from *Saccharomyces cerevisia*. *Anal. Biochem.* 122, , 129-138.

Callen, D. F. (1980) Microbial metabolism of environmental chemicals to mutagens and carcinogens. *Chemical Mutagens: Principles and Methods for Their Detection* 7, 163-188. Eds: deSerres, F. J., Hollaender, A., Plenum Press, New York

Cerniglia, C. E., Gibson, D. T. (1978) Metabolism of naphthalene by cell extracts of *Cunninghamella elegans*. *Arch. Biochem. Biophys.* 186, 1, 121-127.

LeBeault, J. M., Lode, E. T., Coon, M. J. (1978) Fatty acid and hydrocarbon hydroxylation in yeast: Role of cytochrome P-450 in *Candida tropicalis*. *Biochem. Biophys. Res. Commun.* 42, , 413-419.

Wiseman, A. (1980) Xenobiotic-metabolising cytochromes P-450 from microorganisms. *Trends Biochem. Sci.* 5, 4, 102-104.

Wiseman, A., Woods, L. F. J. (1979) Benzo(a)pyrene metabolites formed by the action of yeast cytochrome P-450/P-448. *J. Chem. Tech. Biotechnol.* 29, , 320-324.

Wolf, C. R., Mansuy, D., Nastainczyk, W., Deutschmann, G., Ullrich, V. (1977) The reduction of polyhalogenated methanes by liver microsomal cytochrome P-450. *Mol. Pharmacol.* 13, , 69.

Castro CE, Wade RS, Belser NO (1985) *Biochemistry* 24, 204.

## Microsomal P450

Miller RE, Guengerich FP (1982) Oxidation of trichloroethylene by liver microsomal cytochrome P-450: evidence for chlorine migration in a transition state not involving trichloroethylene oxide. *Biochemistry*, 21, 1090-1097